





Journée Cyanobactéries FIRE 24 avril 2024

Transfer of cyanobacteria and cyanotoxins from fresh to marine waters



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F1 Reservoir water production

F2 St Eloi River

St Floi river

Port

section, M

2-year survey, monthly or bi-weekly, sampling of phytoplanktonic populations, quantification of cyanotoxins microcystins in phytoplankton, in water and in bivalves (LC-MS/MS)

Conclusion

Using sentinel species as bioindicators of cyanotoxins Petit St Gourlais Phytoplankton may not represent water contamination levels Brehot Kerlan Tregen Kerraud La Ville Albertine Le Maguero Spatio-temporal variations in localisation/abundance of **F1** cyanobacteria **Dilution** effect in the estuary **F2** Fleuria Kerlan Sporadic events of cyanotoxin transfer Saint-I Prad Yof Sentinel Yoff species address to changes in cyanotoxin concentrations : filtering bivalves as temporal integrator of cyanotoxin bioavailability **E2** Tremelgon Le Menhi Anodonta anatina sp, (20-30 cm) Bot Bihan Laboratory and multi-site validation Kenva PhD A. Lepoutre 2015-19 Bourg l'Étang *Mytilus edulis* (4-6 cm) M



Cyanobacteria dominance in summer from July to October 2016 and from June to September 2017



Cyanobacteria population dynamics

Conclusion

Recurrent summer proliferation at the freshwater sites in 2016 and 2017 with maximum of **2 millions cells/mL**

Transfer of cyanobacteria to first estuarine site E1 (over 100 000 cells/mL in July/Aug 2017). Progressive dilution

Transfer of cyanobacteria to more saline second estuarine site E2, maximum 1000 cells/mL in Sept-Oct 2016 and June-July 2017 Dilution

> Downstream cell densities related to upstream ones : transfer and not *in situ* growth



26 species of cyanobacteria at the freshwater site F1

Dominance of Microcystis sp (7 species with 3 coexisting)

Some of these cyanobacterial species transfered downstream, with a selection

Bormans, M., Zouher, A., Mineaud, E., Brient, L., Savar, V., Robert, E., Lance, E., 2019. Demonstrated transfer of cyanobacteria and cyanotoxins along a freshwater-marine continuum in France. Harmful Algae



Diversity of cyanobacteria

Conclusion

Microcystis aeruginosa survive despite the fragility of its colonies

Only *P. Agardhii* observed once in the marine section

Progressive species selection Possible species resistance to the longitudinal salinity gradient

Physiological/morphological parameters controlling species selection currently investigated PhD thesis MG Des Aulnois, Ifremer



Frequency of occurrence of intracellular MCs in phytoplanctonic biomass

Max [MC] µg/L Sites FO % intracellular MC phytoplankton (June-November) 2016 2017 F1/F2 100 100 165 **E1** 87 67 1.15 **E2** 38 0.14 17 Μ 0 27 0.03

➢ 9 MC variants (mostly MC-LR, MC-YR, MC-RR)

> Transfer of alive cyanobacterial cells producing MCs, with a progressive decrease of intracellular toxin concentration







Intracellular and extracellular MCs. Bioaccumulated by mussels?



Ability of mussel to integrate MCs between 2 sampling dates and to reveal water contamination levels

			positive samples	
	2016	2017	2016	2017
M. edulis	57 (67)	100 (87)	996,0	152,3
M. edulis	57 (17)	54 (38)	273,7	77,7
M. edulis	20 (0)	27 (27)	14,1	21,3
	M. edulis M. edulis M. edulis	Image: Constraint of the product of	2016 2017 M. edulis 57 (67) 100 (87) M. edulis 57 (17) 54 (38) M. edulis 20 (0) 27 (27)	Image: Provide provide provide same same same same same same same sam



The safe MC threshold concentrations in mussels were evaluated for low (grey full dash) or high (grey dotted dash) consumers based on the WHO TDI of 40 ng MCs/day/kg, and for low (black full dash) or high (black dotted dash) consumers based on the new French guideline value of ANSES of 1 ng MCs/day/kg BW.

Lance, E., Lepoutre, A., Savar, V., Robert, E., Bormans, M., Zouher, A., 2021. *In situ* use of bivalves and passive samplers to reveal water contamination by microcystins along a freshwater-marine continuum in France. Water Research







Interest of bivalves as bioindicators of MCs, risk for human health

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AND THANK YOU FOR YOUR ATTENTION!

